

# *FMDV as a model for lethal mutagenesis*

*Esteban Domingo*

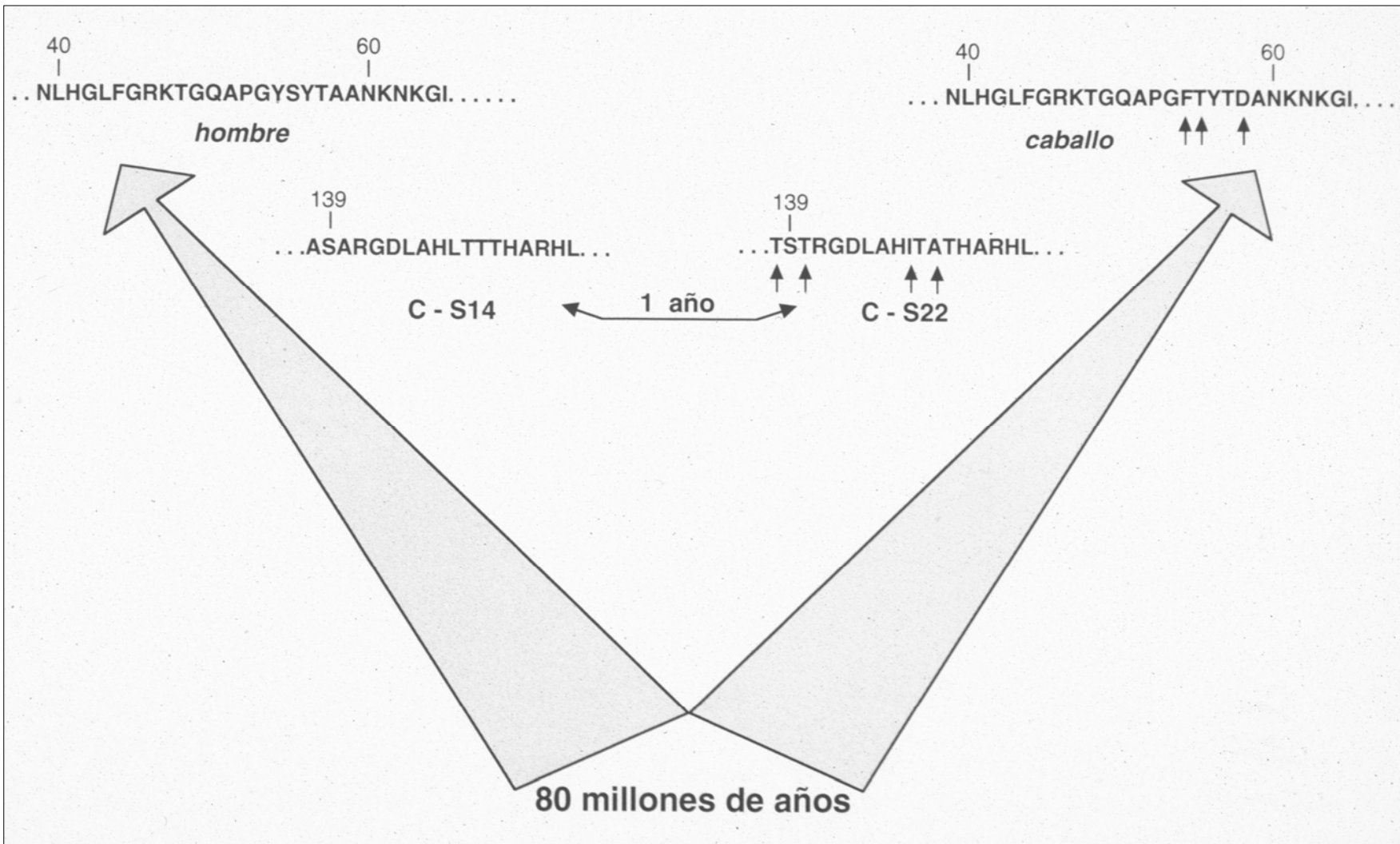
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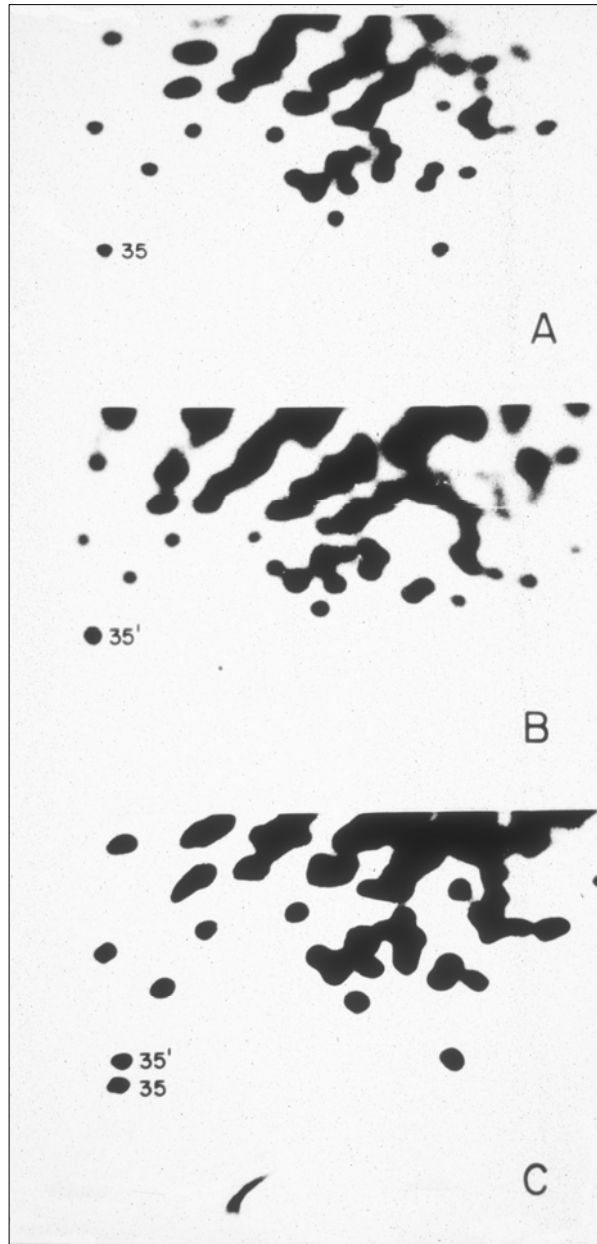
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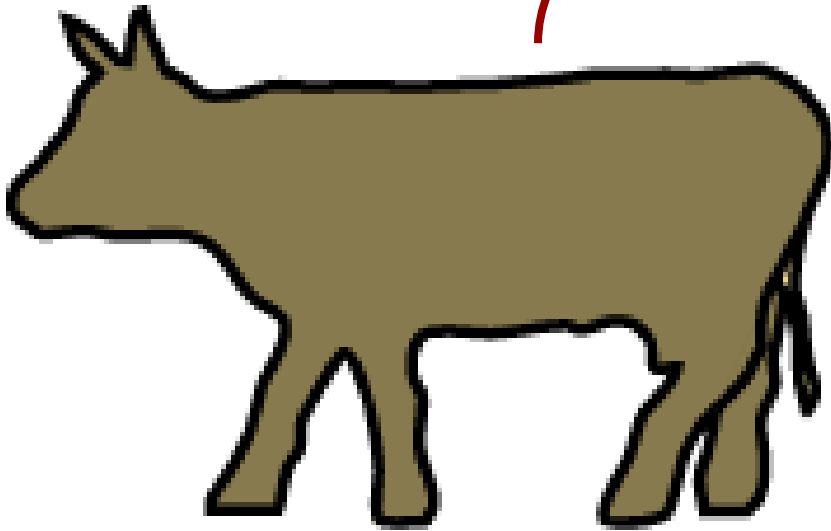
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1. *Some historical slides, seen 28 years later*
2. *High error rates and virus population dynamics demand new antiviral approaches*
3. *How FMDV has helped in the current development of lethal mutagenesis*
4. *Prospects. Links with anti-cancer therapy*

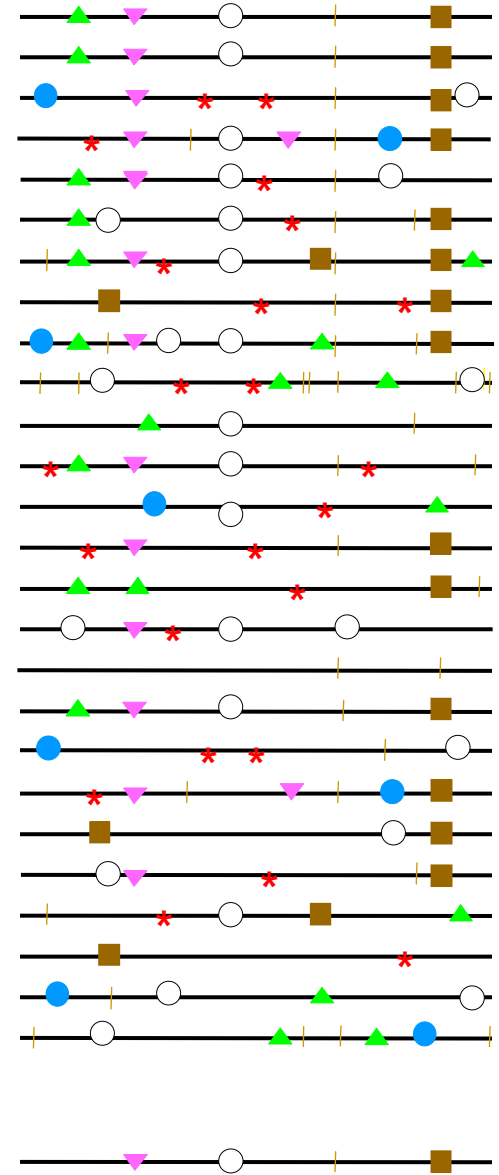
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A99m	C	S	C	G	G	U	A	U	A	B	C	U	A	C	U	B	A	U	S	CC	CC	U	U	A	U	
A10	U	U	C	U	G	U	U	U	U	C	U	U	A	A	U	U	U	U	U	U	U	U	U	U	U	U
A22	C	C	C	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U
A24	A	M	M	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
C88	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U
C819	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U
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C3P8	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U
D18r	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U
D18p	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U
D187A	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U
D1C	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U	U
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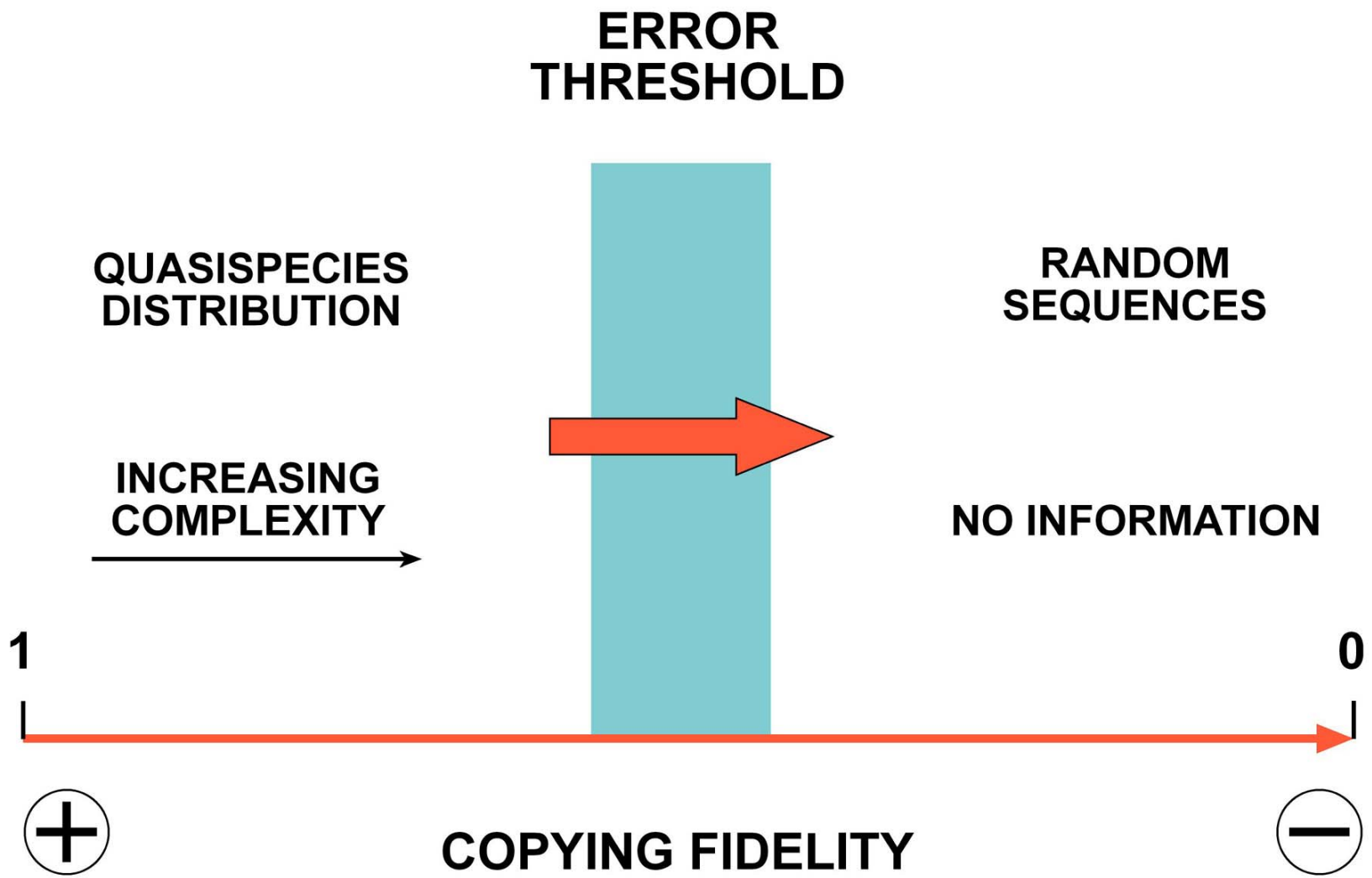




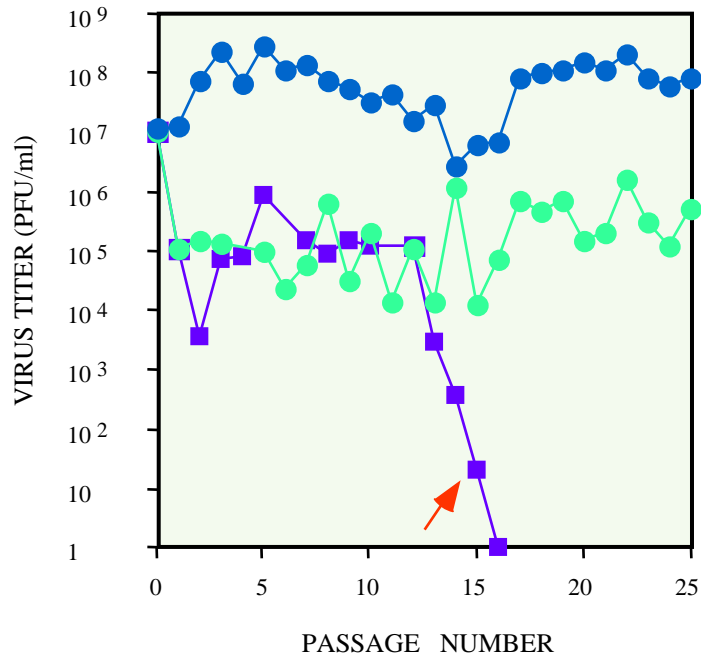
MUTANT SPECTRUM



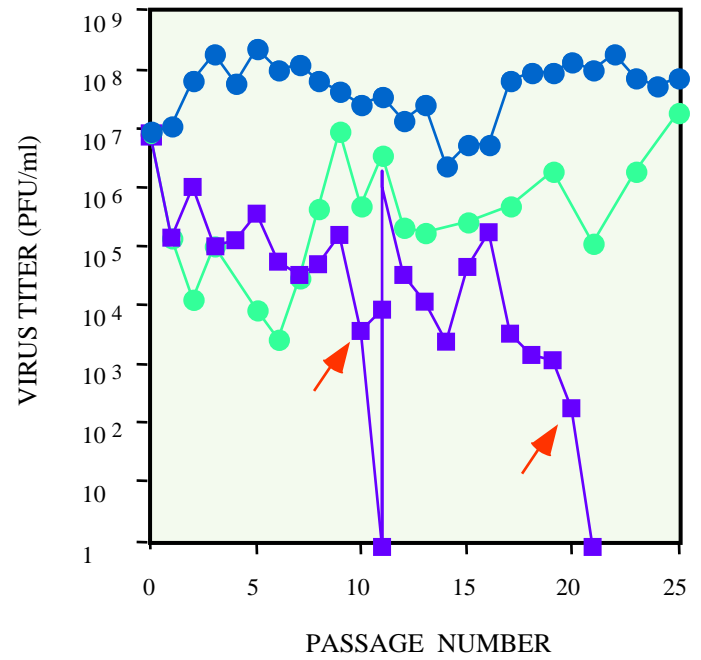
CONSENSUS



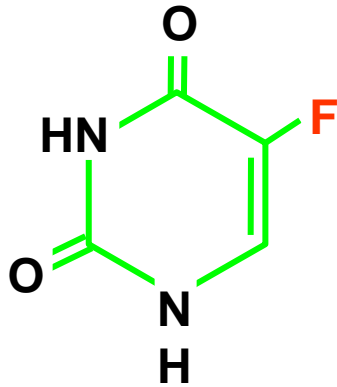
# FU



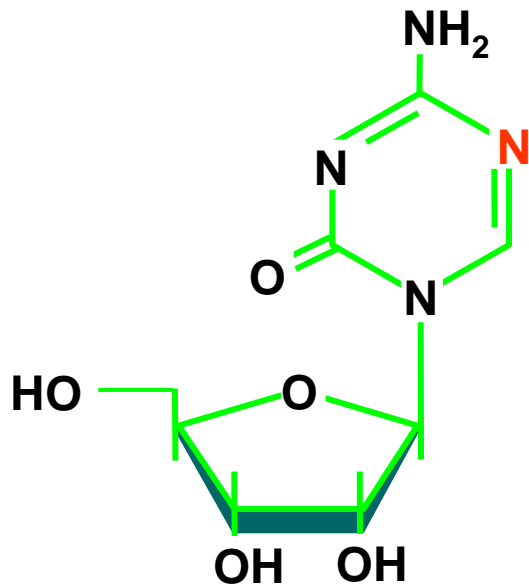
# AZC



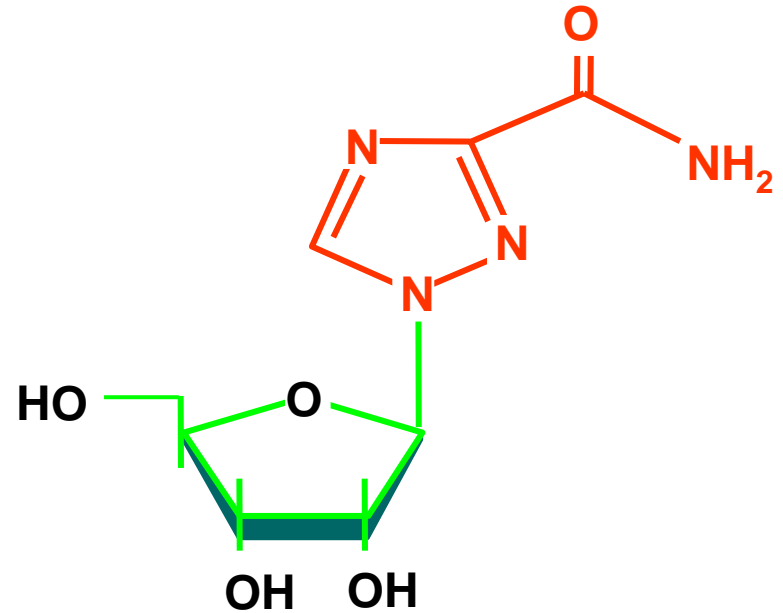




**5 - Fluorouracil**

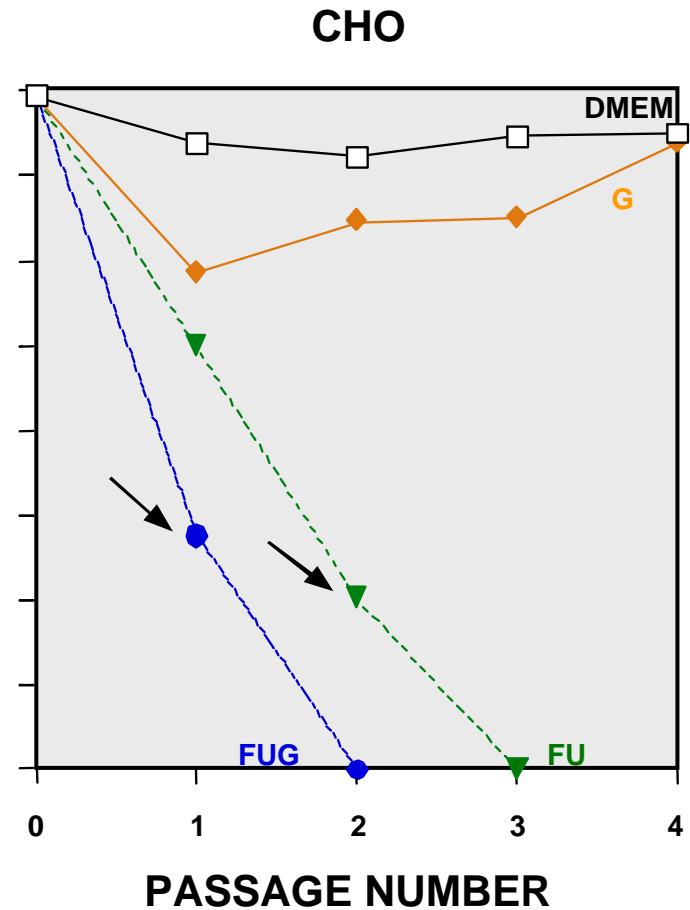
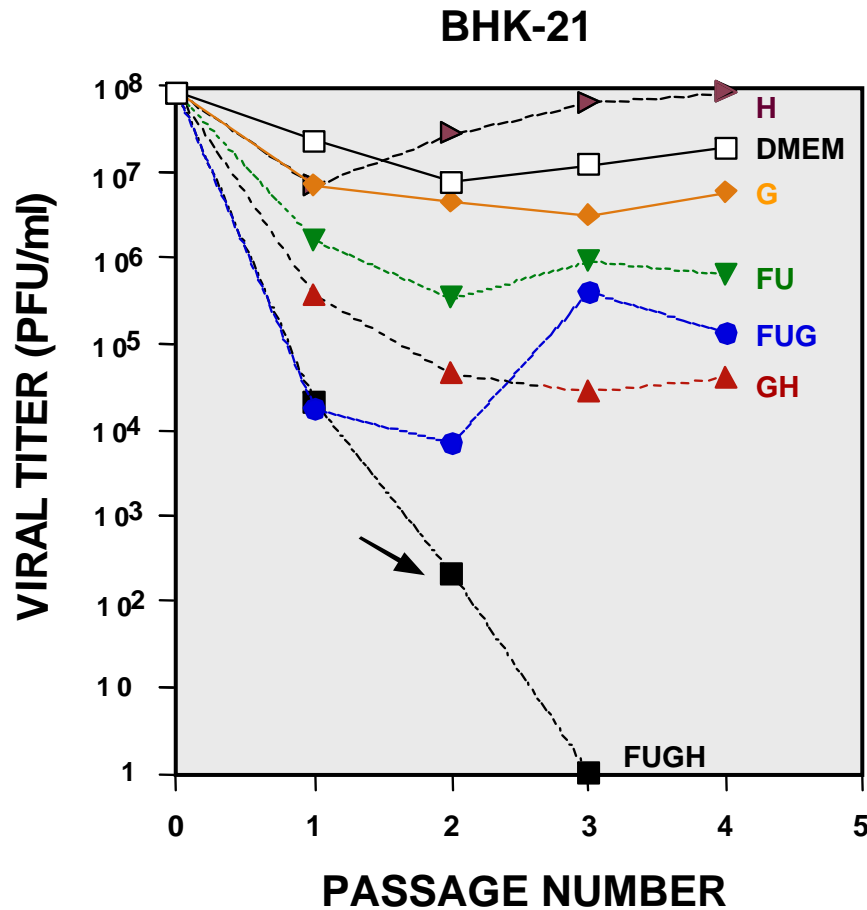


**5 - Azacytidine**



**Ribavirin**

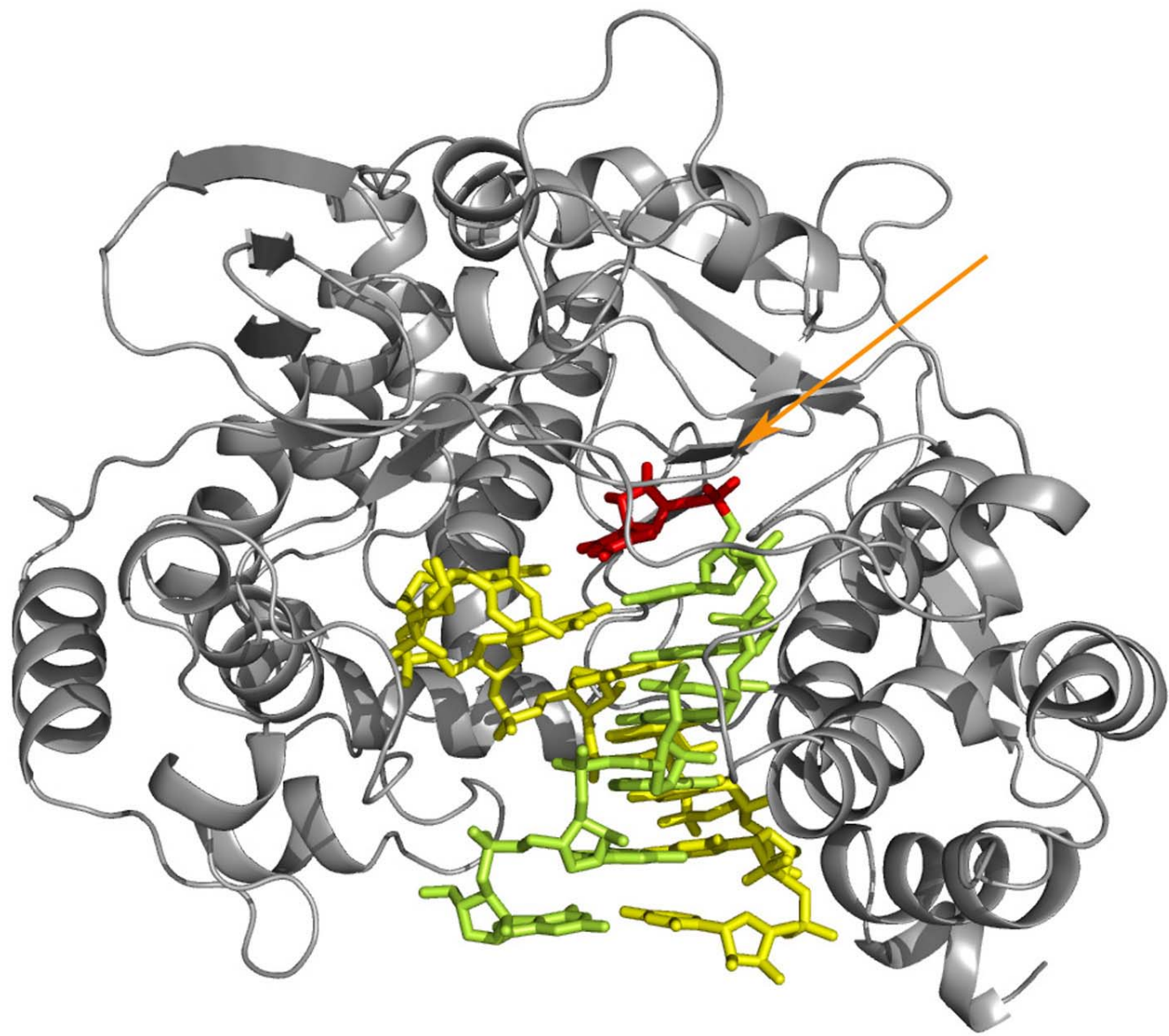
# FMDV MARLS (RELATIVE FITNESS 130 IN BHK-21 CELLS)

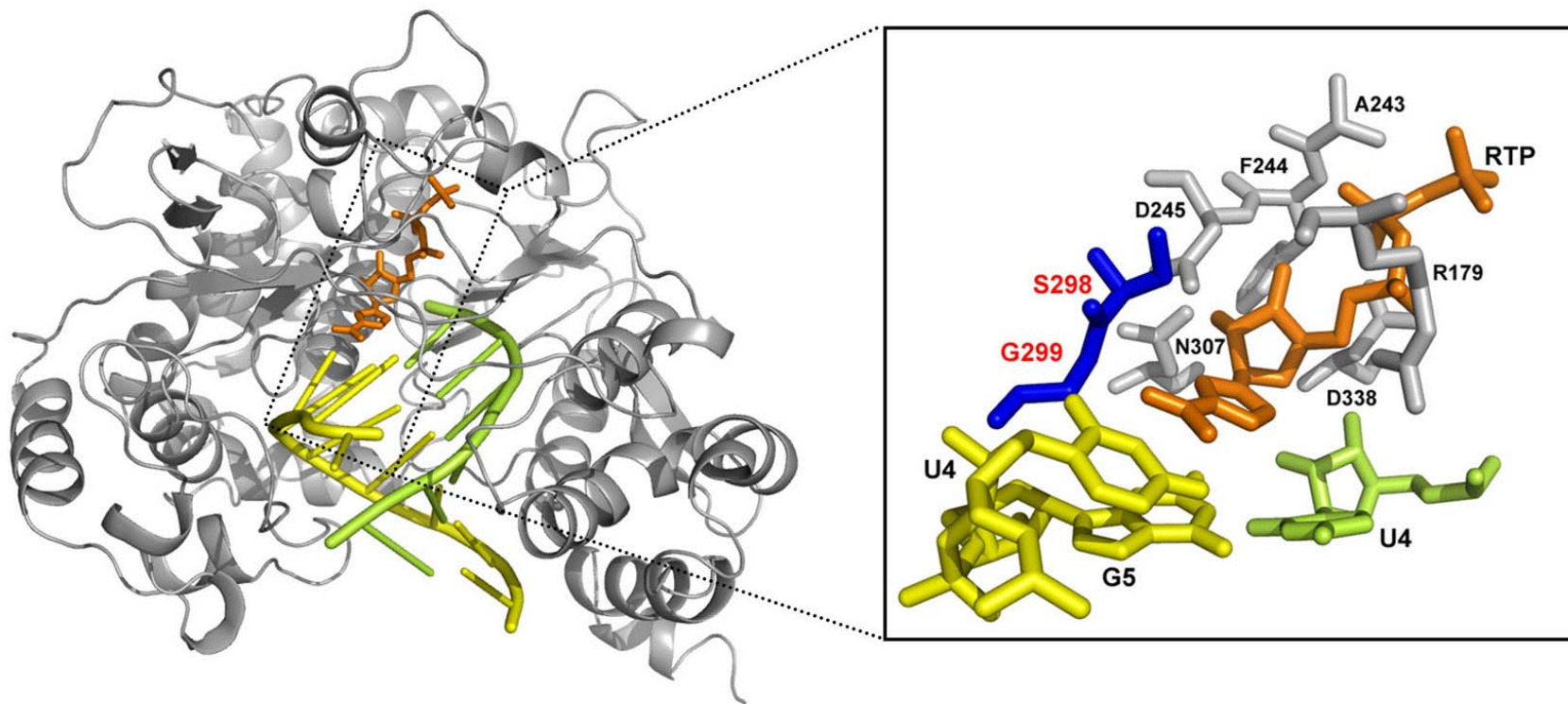


- *Preextinction FMDV RNA interferes with infectious FMDV RNA (despite low relative fitness of preextinction FMDV:  $\leq 0,183$  relative to parental virus, fitness 1)*
- *The transition of FMDV towards extinction is accompanied of:*
  - *Increases of 2.0- to 11.1- fold in mutation frequency (measured with components of mutant spectra)*
  - *Decreases of about  $10^3$ -fold in specific infectivity (infectious RNA/total RNA  $\sim 2.5 \times 10^{-7}$ )*
  - *Invariant consensus nucleotide sequence*
- *The same initial inhibitory activity but without a mutagenic agent does not lead to extinction*

# *Molecular basis of mutagenesis*

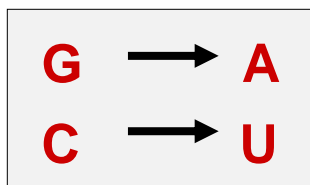
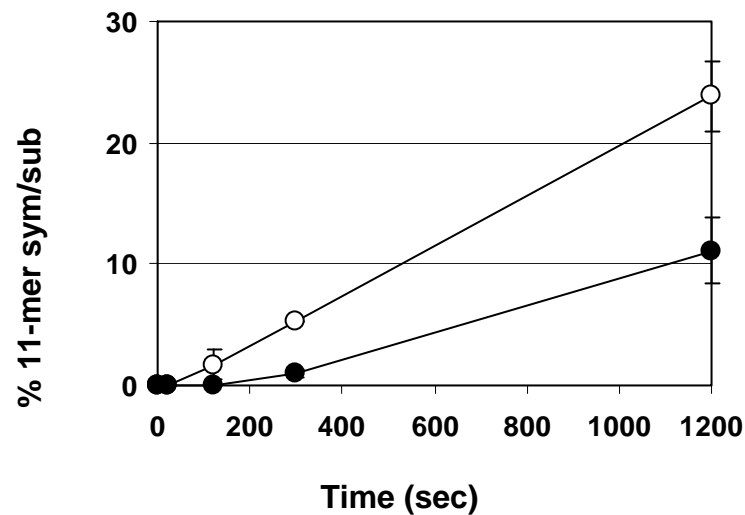
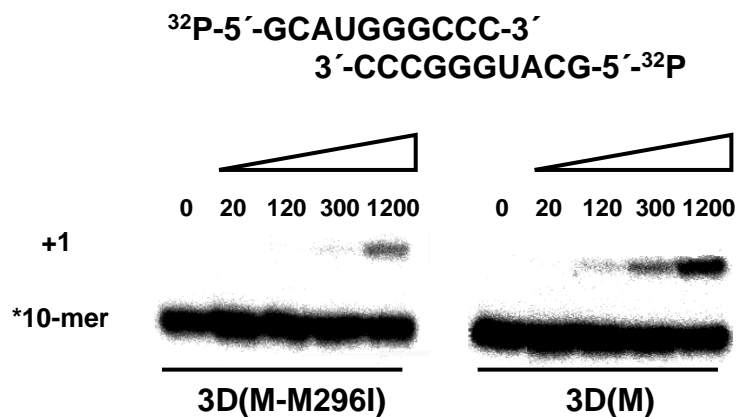
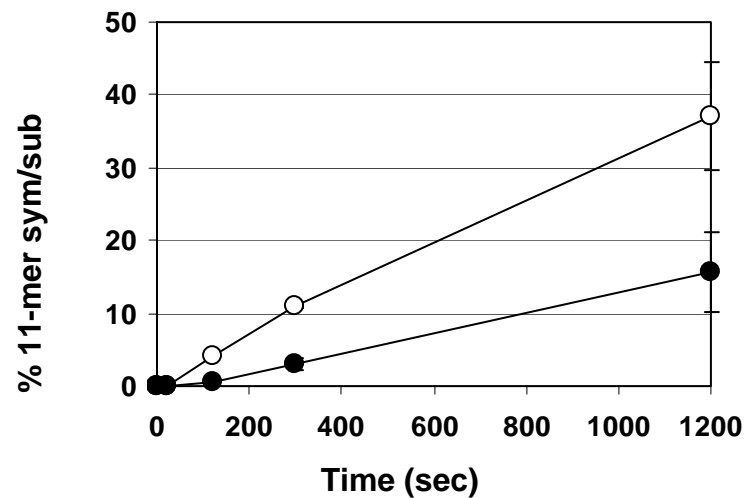
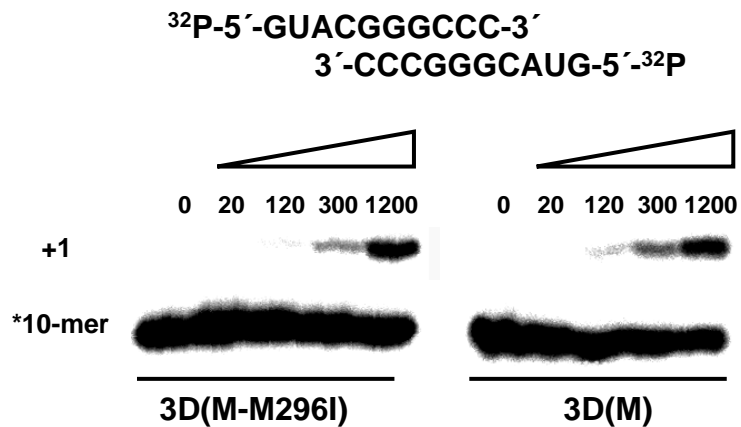
- *Base and nucleoside analogues are converted to their nucleotides which can be active in mutagenesis by at least two mechanisms:*
- *The nucleoside-triphosphates can be incorporated by the viral polymerases and induce transition mutations*
- *Nucleotide analogues may inhibit enzymes of nucleotide metabolism thereby altering intracellular nucleotide pools*





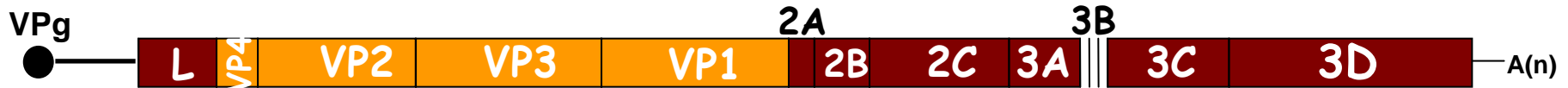
## *FMDV 3D mutant M296I selected by ribavirin*

- *The substitution increases viral fitness in the presence of ribavirin, but not in the absence of ribavirin*
- *The mutant 3D shows decreased capacity to incorporate ribavirin monophosphate in the place of GTP or ATP*
- *In other biochemical reactions (standard polymerization assays, VPg uridylylation, RNA binding) the mutant 3D behaves as the wild type 3D*





# FMDV CAPSID MUTANTS



## Cápsid

Q2027A

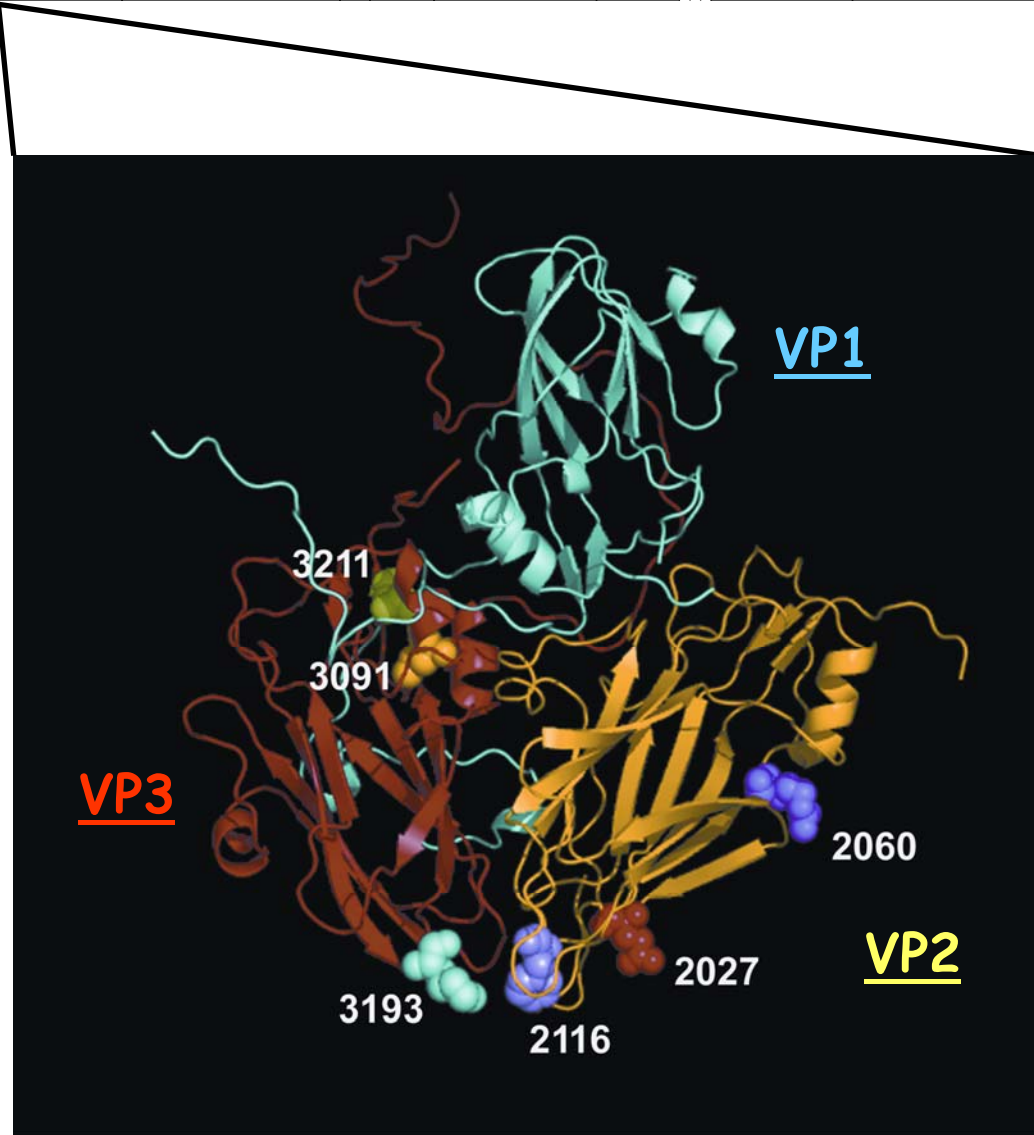
R2060A

F2116A

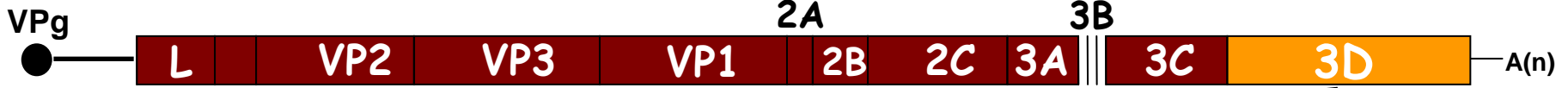
K3193A

L3091R

L3211P



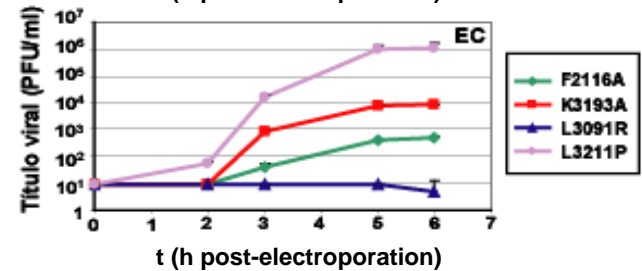
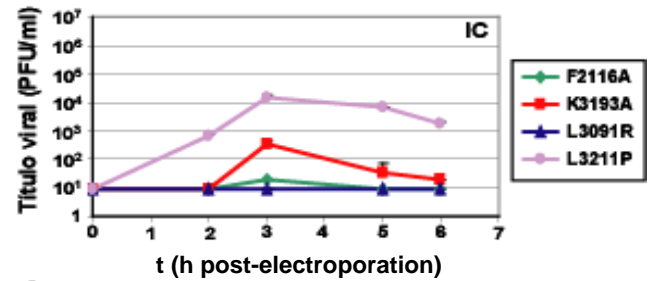
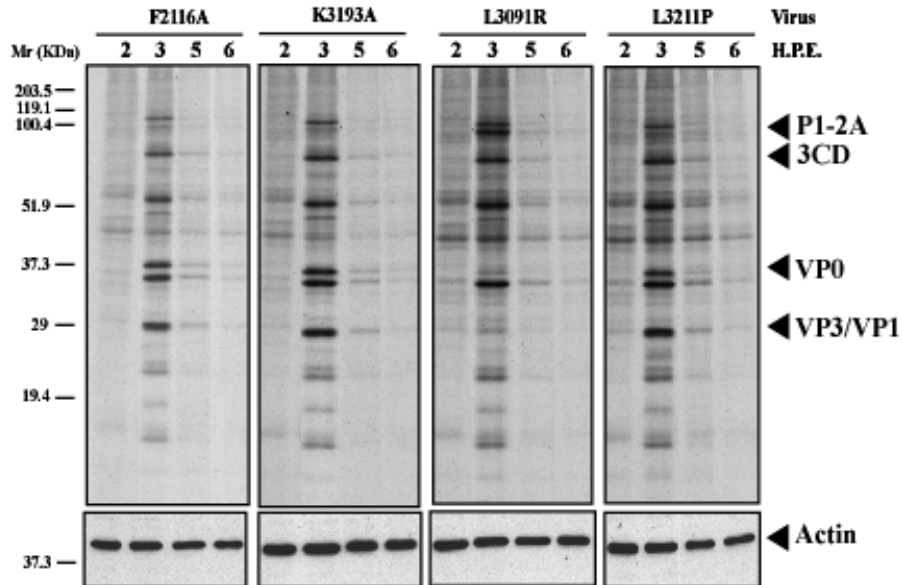
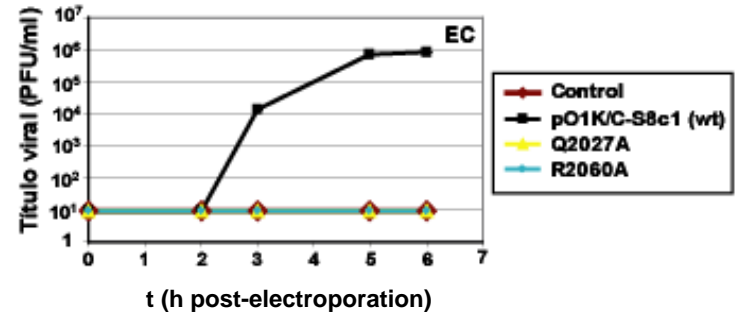
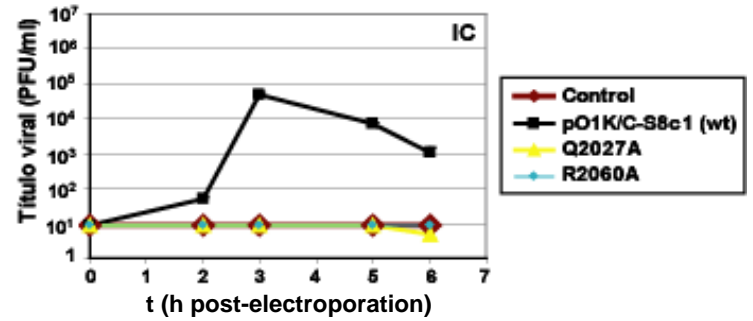
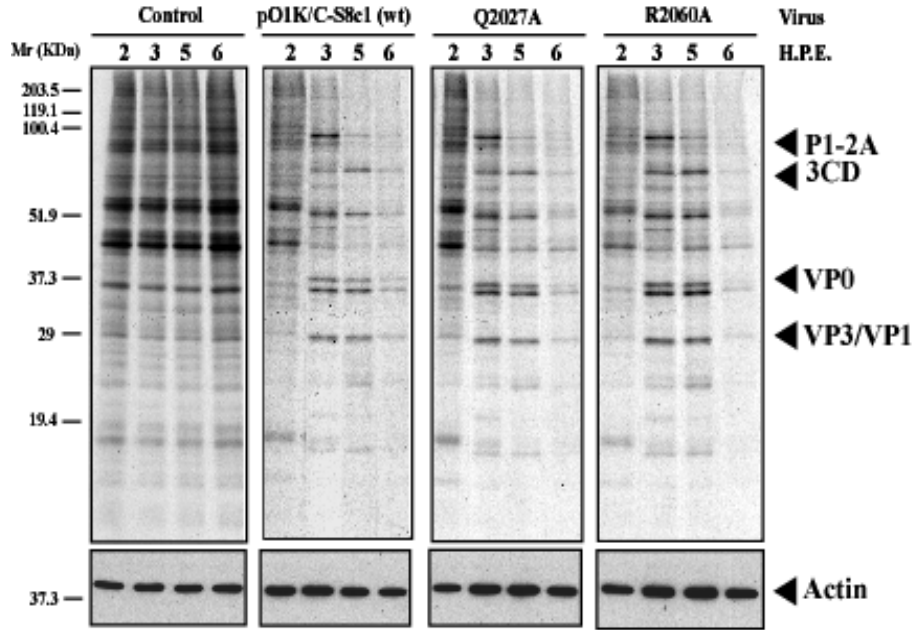
# FMDV POLYMERASE MUTANTS



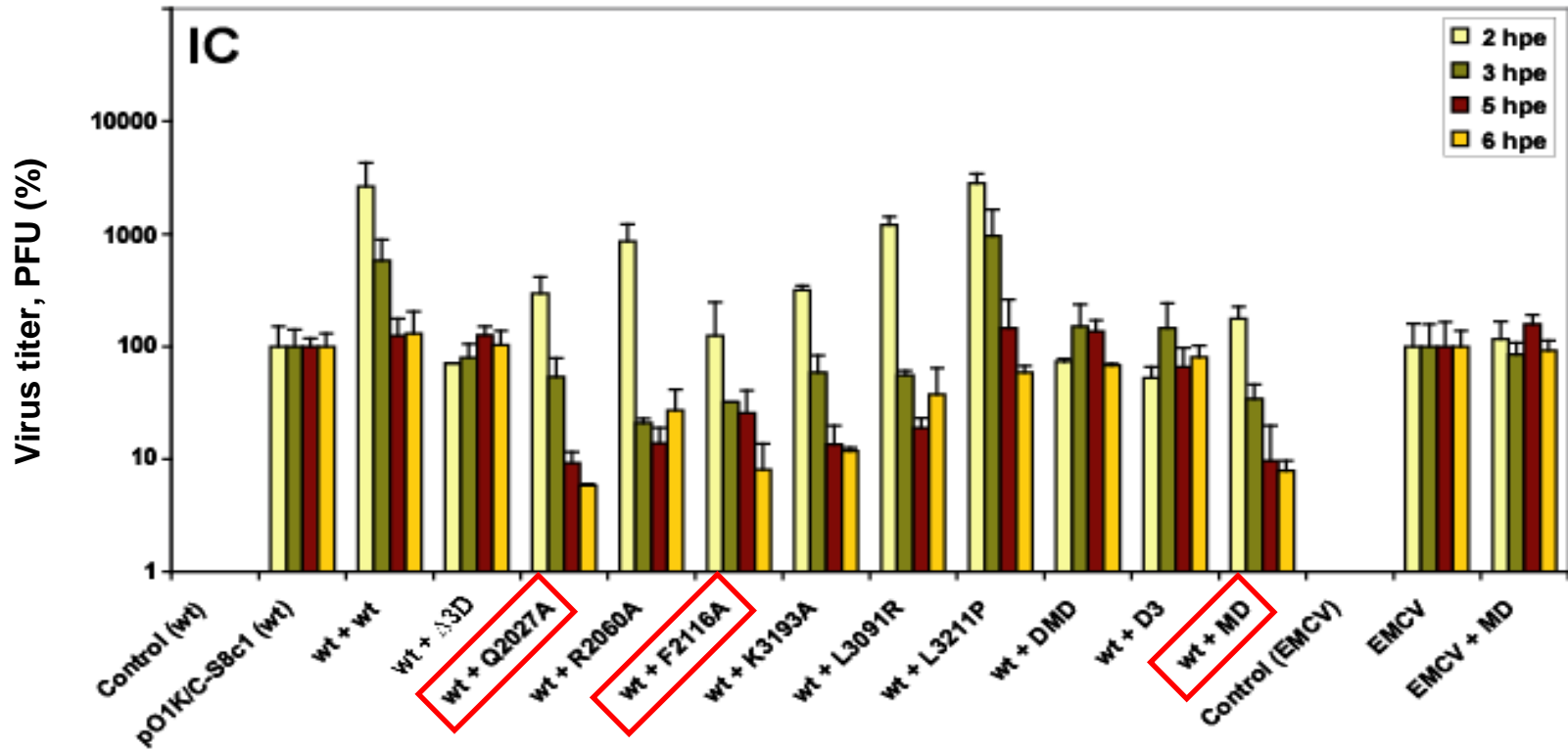
## Polymerase

- |     |   |       |
|-----|---|-------|
| DMD | { | G118D |
|     |   | V239M |
|     |   | G373D |
| D3  | { | D338A |
| MD  | { | V239M |
|     |   | G373D |

# CAPSID MUTANTS OF FMDV



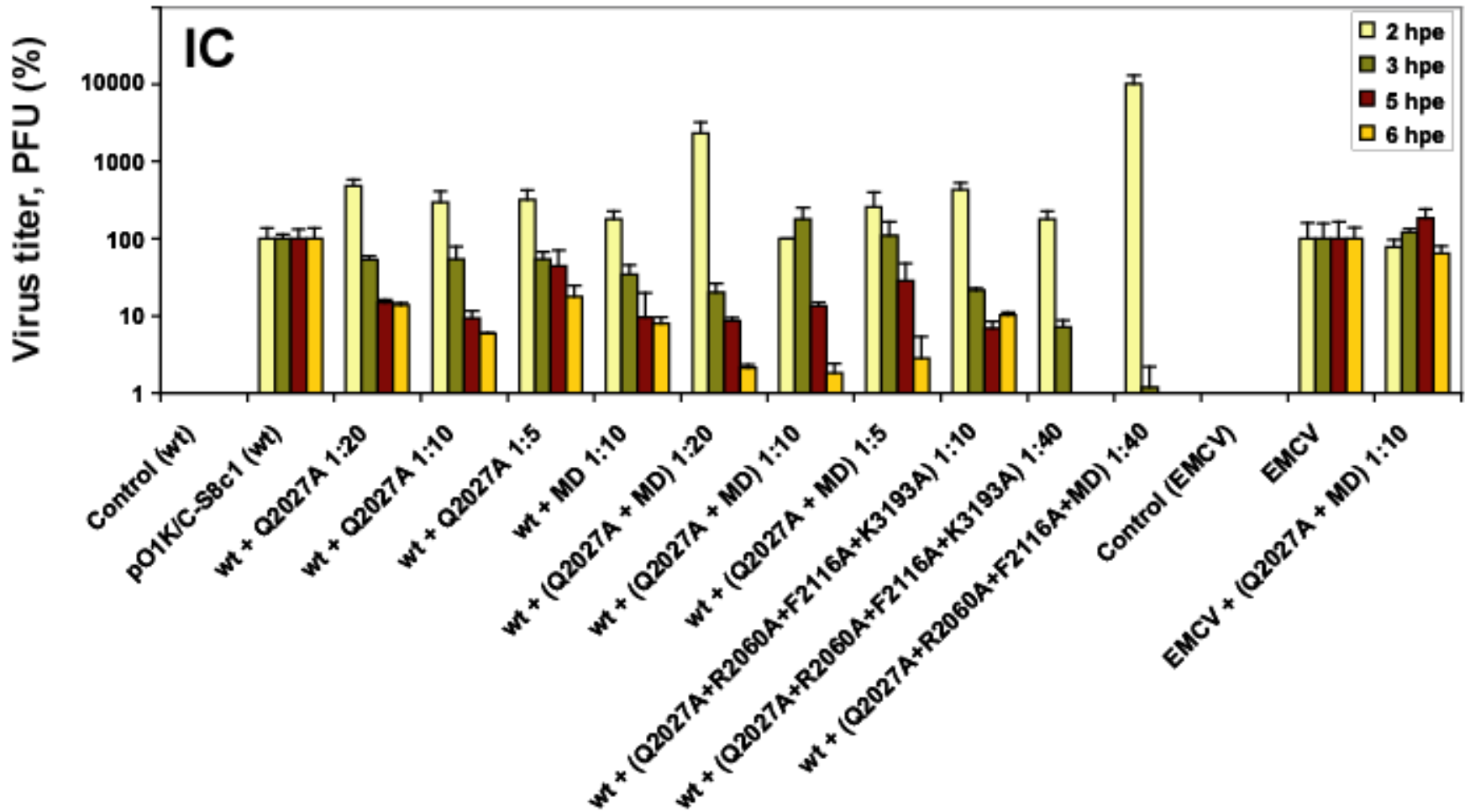
# INTERFERENCE BY INDIVIDUAL FMDV MUTANTS



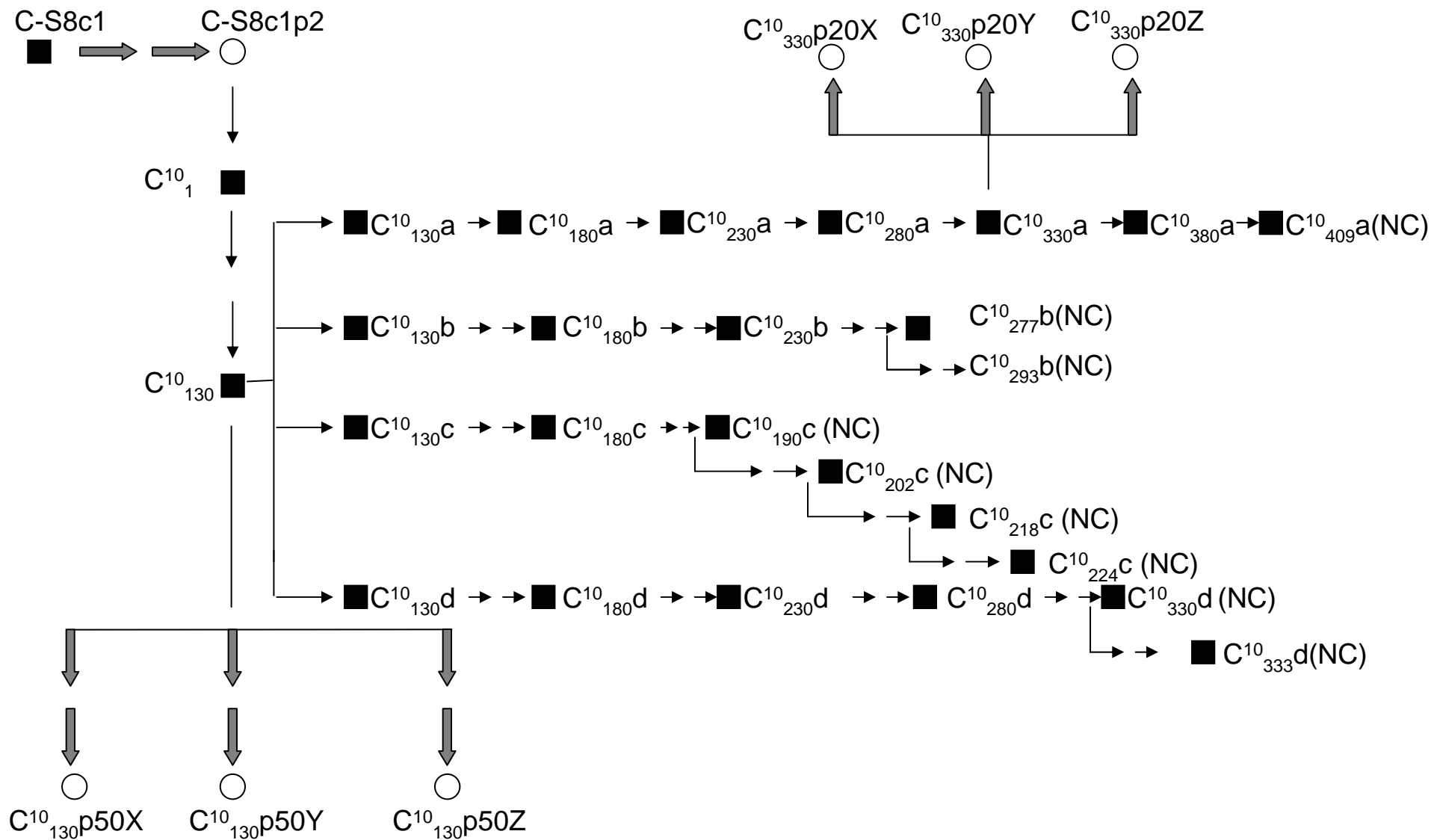
**SUMMARY OF REPLICATIVE AND INTERFERENCE**  
**PROPERTIES OF FMDV MUTANTS**

Virus	Amino acid substitution	Protein	Intracellular replication	Infectious progeny		Interference
				Intracellular	Extracellular	
pO <sub>1</sub> K/C-S8c1 or pMT28	—	—	++	++	++	—
<b>Q2027A</b>	Q27A	VP2	++	—	—	<b>++</b>
<b>R2060A</b>	R60A	VP2	++	—	—	+
<b>F2116A</b>	F116A	VP2	++	—	—	<b>++</b>
<b>K3193A</b>	K193A	VP3	++	+/-	+/-	+
<b>L3091R</b>	L91R	VP3	++	—	—	+
<b>L3211P</b>	L211P	VP3	++	++	++	—
<b>DMD</b>	G118D					
	V239M	3D	—	—	—	—
	G373D					
<b>D3</b>	D338A	3D	—	—	—	—
<b>MD</b>	V239M	3D	++	+	+	<b>++</b>
	G373D					

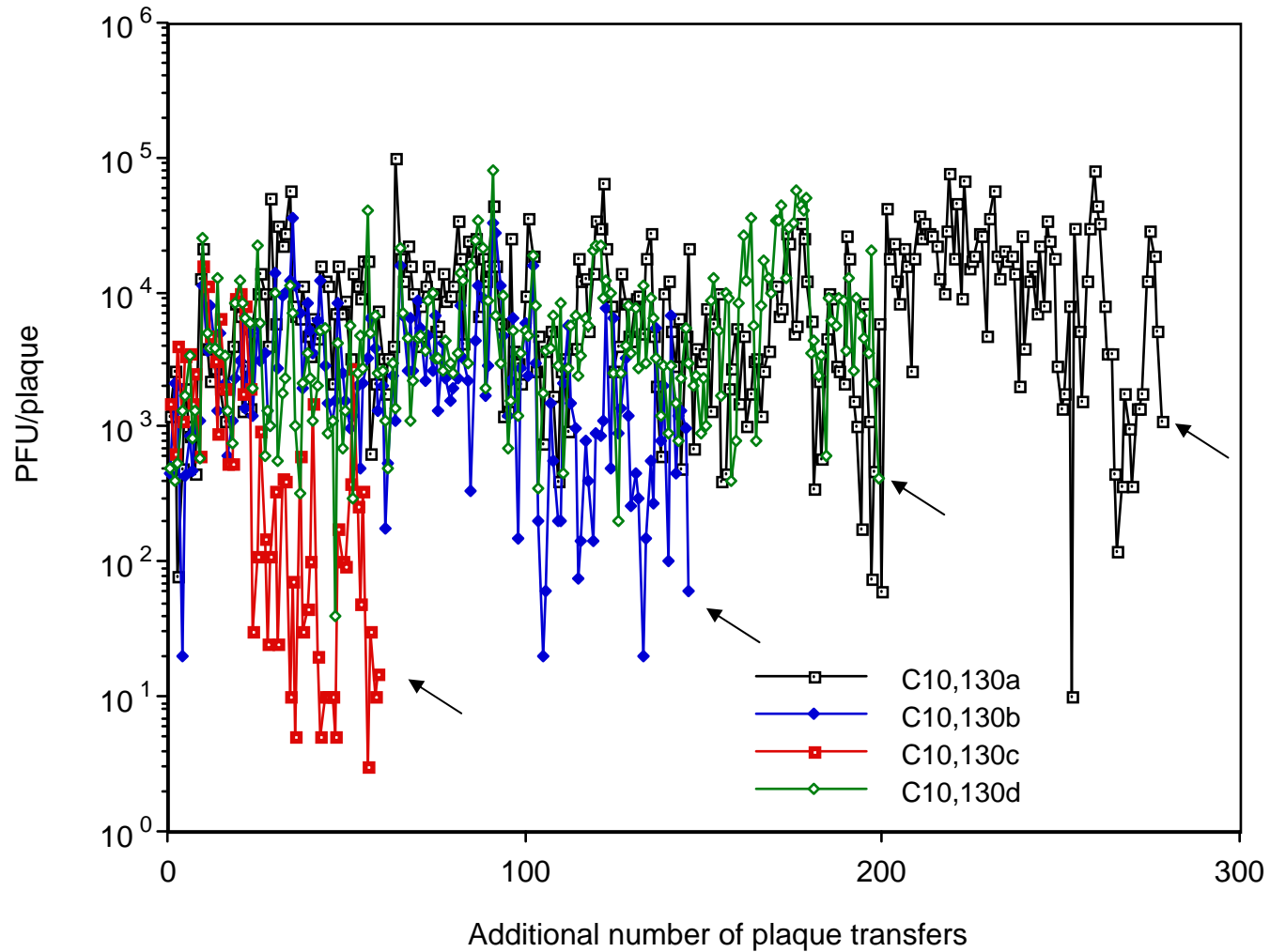
# INTERFERENCE BY MIXTURES OF FMDV MUTANTS



- *Specific mutants can interfere with FMDV replication*
- *Some lethal mutants did not interfere, while some replication-competent mutants (notably polymerase mutant MD) exerted a strong interference*
- *Interference was stronger with combinations of capsid and polymerase mutants*



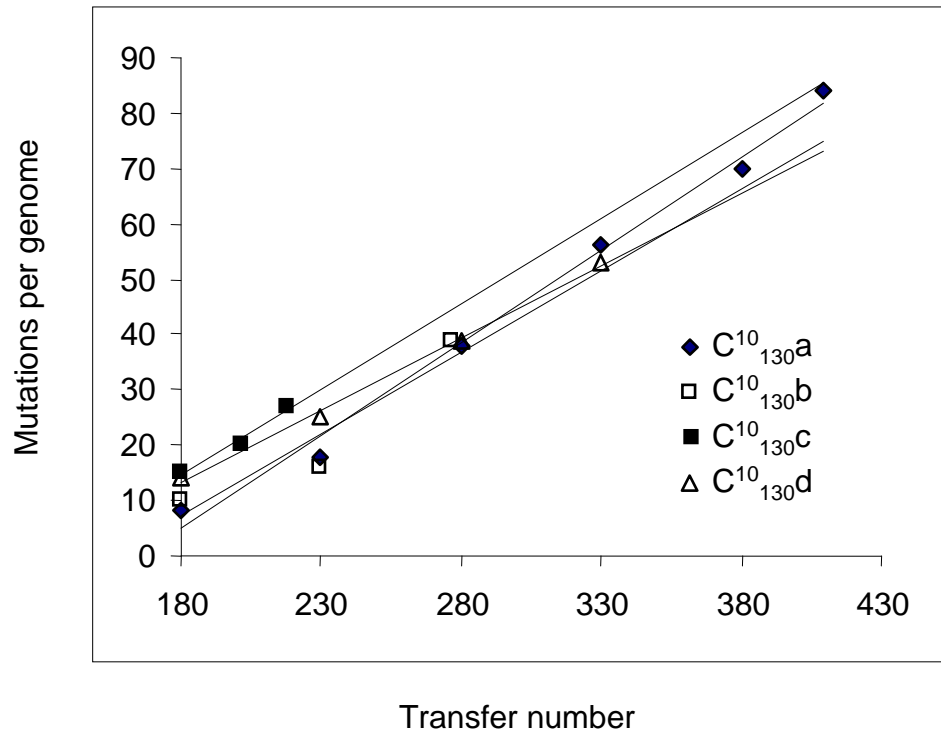




## *Specific infectivity*

FMDV	RNA molecules in plaque	PFU in plaque	Specific infectivity PFU / RNA
C <sup>10</sup> <sub>1</sub>	4.5 x 10 <sup>8</sup>	1.1 x 10 <sup>5</sup>	2.4 x 10 <sup>-4</sup>
NC clones (218-409 transfers)	3.0 x 10 <sup>6</sup> 3.8 x 10 <sup>6</sup>	< 5	< 1.7 x 10 <sup>-6</sup> < 1.3 x 10 <sup>-6</sup>

- *NC clones establish a persistent infection in BHK-21 cells, without a phase of cell killing*



## *Plaque – to- plaque transfers*

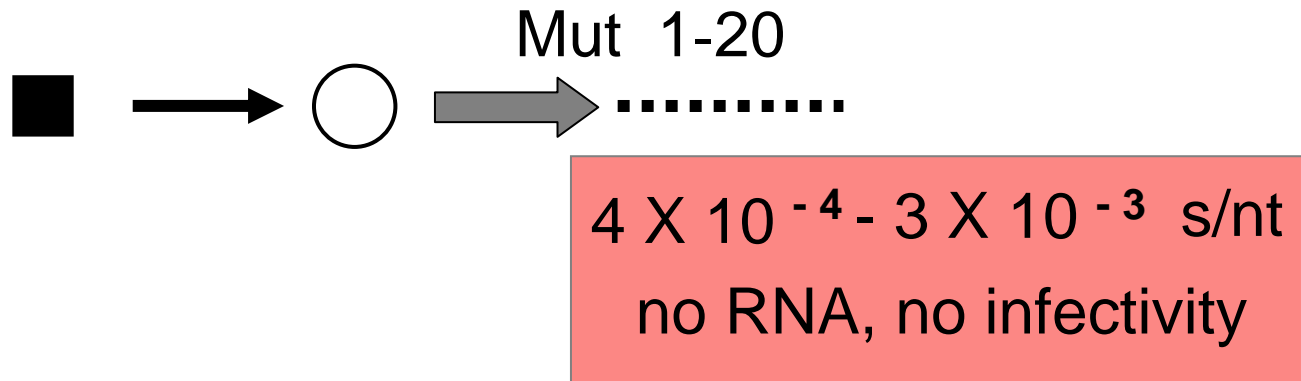
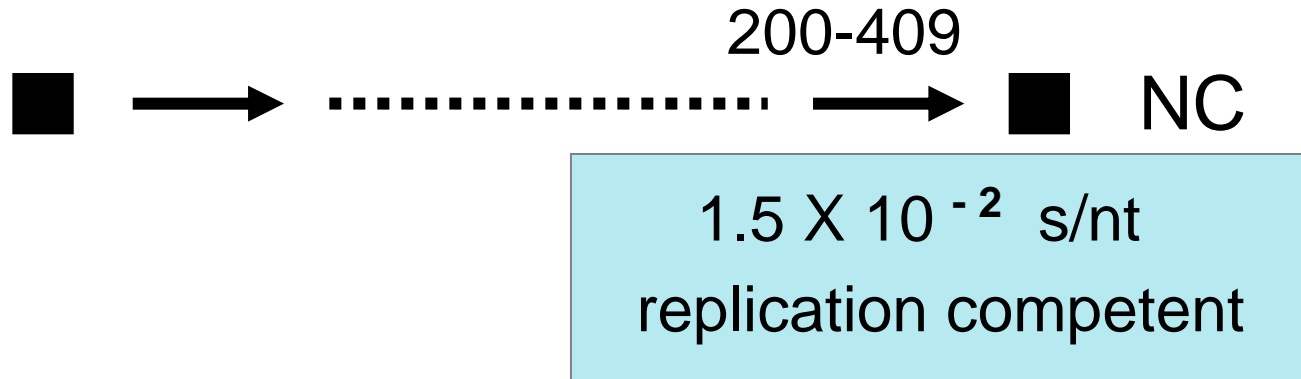
*0.26 – 0.34 mutations per genome per transfer*

$$dn / ds = 0.13 – 0.16$$

## *Large population passages*

*0.10 – 0.25 mutations per genome per passage*

$$dn / ds = 0.50 – 1.20$$



## *Our current view of intra-quasispecies interactions and lethal mutagenesis*

- *Positive interactions (complementation) and negative interactions (interference) occur within the viral quasispecies. In well adapted mutant distributions, complementation dominates*
- *With enhanced mutagenesis, negative interactions (interference) operate, and contribute to a decrease of infectivity. Infectivity loss precedes replication collapse, probably due to the involvement of more functions in infectivity than in genome replication*
- *As mutagenesis progresses, the proportion of genomes with interfering mutations and lethal mutations increases, leading eventually to the complete replicative collapse and virus extinction*

***•The transition into error catastrophe does not occur through “evaporation” into the entire sequence space. This is physically impossible. A “phenotypic” or “extinction” threshold intervenes prior to the classical genotypic “error” threshold. This is an obvious extension of “error catastrophe” theory to real viruses***

# *Prospects of lethal mutagenesis as an antiviral therapy*

- *An increasing number of specific mutagenic nucleoside analogues are under study. Links with anti-cancer chemotherapy*
- *A clinical trial (phase 1b) was initiated in 2005 with about 40 HIV-1-infected patients who failed HAART. Phase 2 scheduled for 2007. It involves administration of a new nucleoside analogue KP-1461 (Koronis Pharmaceuticals Inc., Redmond, WA, USA)*



## References

- *Sierra et al. (2000) J. Virol 74: 8316-8323*
- *Ferrer-Orta et al. (2007) Proc. Natl. Acad. Sci. 104: 9463-9468*
- *Perales et al. (2007) J. Mol. Biol. 369: 985-1000*
- *Escarmís et al. (2008) J. Mol. Biol. 367: 367-379*

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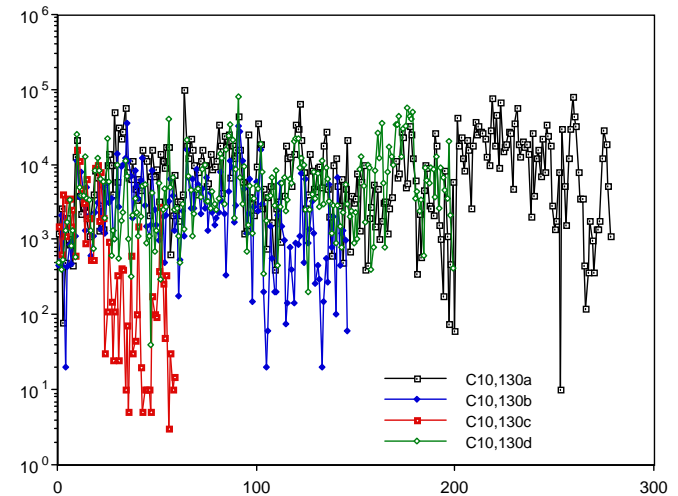
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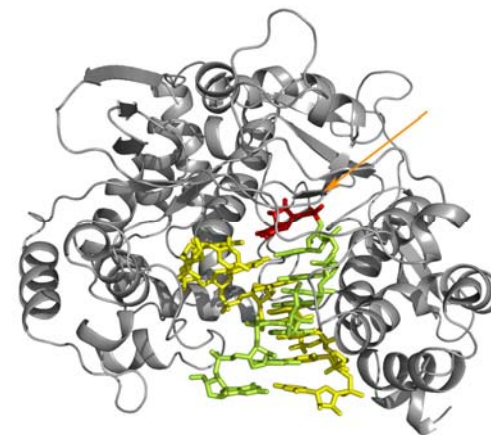
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